



Europäisches
Patentamt

European
Patent Office

Office européen
des brevets

12. 03. 04

RECEIVED

24 MAR 2004

WIPO

PCT

Bescheinigung

Certificate

Attestation

Die angehefteten Unterla-
gen stimmen mit der
ursprünglich eingereichten
Fassung der auf dem näch-
sten Blatt bezeichneten
europäischen Patentanmel-
dung überein.

The attached documents
are exact copies of the
European patent application
described on the following
page, as originally filed.

Les documents fixés à
cette attestation sont
conformes à la version
initialement déposée de
la demande de brevet
européen spécifiée à la
page suivante.

Patentanmeldung Nr. Patent application No. Demande de brevet n°

03075928.6

**PRIORITY
DOCUMENT**
SUBMITTED OR TRANSMITTED IN
COMPLIANCE WITH RULE 17.1(a) OR (b)

Der Präsident des Europäischen Patentamts;
Im Auftrag

For the President of the European Patent Office

Le Président de l'Office européen des brevets
p.o.

R C van Dijk



Anmeldung Nr:
Application no.: 03075928.6
Demande no:

Anmeldetag:
Date of filing: 31.03.03
Date de dépôt:

Anmelder/Applicant(s)/Demandeur(s):

Stichting
Researchfonds Pathologie
Jekerstraat 19 2HG
1078 LW Amsterdam
PAYS-BAS

Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
If no title is shown please refer to the description.
Si aucun titre n'est indiqué se référer à la description.)

Detection of HPV-induced invasive cancers and their precursor lesions with
invasive potential

In Anspruch genommene Priorität(en) / Priority(ies) claimed / Priorité(s)
revendiquée(s)
Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/
Classification internationale des brevets:

C12Q1/68

Am Anmeldetag benannte Vertragstaaten/Contracting states designated at date of
filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HU IE IT LU MC NL
PT SE SI SK TR LI

31. 03. 2003

P63729EP00

(83)

Title: Detection of HPV-induced invasive cancers and their precursor lesions with invasive potential

FIELD OF THE INVENTION

The invention is in the field of medicine and is concerned with a molecular diagnostic marker for human papillomavirus (HPV)-induced invasive cancers and their precursor lesions such as invasive cervical cancer, premalignant cervical lesions with invasive potential and high-risk human papillomavirus (HPV)-induced non-cervical invasive cancers. In particular the present invention relates to the use of the *TSLC1* gene as marker for HPV-induced invasive cancers and their precursor lesions.

10 BACKGROUND OF THE INVENTION

Cancer of the uterine cervix is the second most common cancer in women world-wide and is responsible for approximately 250.000 cancer deaths a year.

15 Cervical cancer development is characterized by a sequence of premalignant lesions, so called cervical intraepithelial neoplasia (CIN), which are graded I to III, referring to mild dysplasia (CIN I), moderate dysplasia (CIN II) and severe dysplasia/carcinoma *in situ* (CIN III).

Over the past decade it has been well established that cervical carcinogenesis is initiated by an infection with high-risk human papillomavirus (HPV). Expression of the viral oncogenes E6 and E7, which disturb the p53 and Rb tumor suppressor pathways, respectively, has been shown to be essential for both the onset of oncogenesis and the maintenance of a malignant phenotype. However, consistent with a multistep process of carcinogenesis, additional alterations in the host cell genome are required for progression of an hr-HPV infected cell to an invasive carcinoma.

20

25

In line with multiple events underlying cervical carcinogenesis is the observation that only a small proportion of women infected with high-risk HPV will develop high-grade premalignant cervical lesions (CIN III) or cervical cancer, and in most women with premalignant cervical lesions the lesions regress spontaneously.

However, at present no markers exist to predict which premalignant lesions will regress or ultimately progress to cervical cancer. Therefore, general medical practice comprises the treatment of all women with morphologically confirmed CIN II and CIN III, in order to prevent the development of cervical cancer. Consequently, many women are unnecessarily treated and unnecessarily worried.

SUMMARY OF THE INVENTION

It has now surprisingly been found that the tumor suppressor lung cancer 1 gene (*TSLC1*) is involved as a tumor suppressor gene in cervical carcinogenesis and that *TSLC1* silencing, or a low level of expression of the *TSLC1* gene, is an important determinant of cervical carcinogenesis. Compared to its role in the development of a subset of lung carcinomas, the silencing of *TSLC1* plays an even more predominant role in cervical carcinogenesis. *TSLC1* and the gene products thereof thus provide valuable molecular markers to diagnose invasive cervical lesions and/or to (better) predict which premalignant lesions have a high chance of progressing to cervical cancer.

Cervical cancer is almost exclusively associated with human papillomavirus (HPV) infection. Human papillomaviruses, constitute a group of more than 100 types of viruses, as identified by variations in DNA sequence. The various HPVs cause a variety of cutaneous and mucosal diseases. Certain types may cause warts, or papillomas, which are benign (noncancerous) tumors. Others have been found to cause invasive carcinoma of the uterine cervix.

HPVs are broadly classified into low-risk and high-risk types, based on their ability to induce malignant changes in infected cells. Low risk HPV types such as 1, 2, 4, 6, 11, 13 and 32 are primarily associated with benign lesions or common warts while the high risk types, such as 16, 18, 31, 33, 35, 5 39, 45, 51, 52, 56, 58, 59, 68, 73 and 82 are primarily associated with premalignant and malignant epithelial lesions. These high-risk types of HPV cause growths that are usually flat and nearly invisible, as compared with the warts caused by low-risk types, e.g. HPV-6 and HPV-11.

Therefore, the present invention is not only suited to detect invasive 10 cervical cancer associated with tumor suppressor lung cancer 1 (TSLC1), but also other invasive cancers that are induced by HPV, particularly of the high-risk type and provides a method for the risk assessment of premalignant lesions becoming invasive.

Accordingly, the present invention provides methods of detecting 15 HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1) in a subject in need thereof, said method comprising contacting a cell component of a test cell of the subject with a reagent that detects the level of the cell component in the test cell and determining a modification in the level of the cell component in the test cell as 20 compared with a comparable healthy cell, wherein the cell component indicates the level of TSLC1 in the cell and the modification indicates the presence of HPV-induced invasive cancer.

Very suitable HPV-induced invasive cancers or precursor lesions thereof in the context of the present invention are invasive cervical cancers 25 and premalignant cervical lesions with invasive potential, but also invasive cancers and premalignant lesions that are induced by HPV in other tissues such as from oral cavity, oropharynx, anus, rectum, penis, vulva, etc.

A test cell may be a neoplastic cell, a proliferating cervical cell, or any other cell wherein the presence of an HPV-induced invasive cancer or

precursor lesion thereof associated with tumor suppressor lung cancer 1 is to be detected.

In another embodiment, the present invention provides methods of detecting HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1) in a subject in need thereof, said method comprising contacting a target cellular component of a test cell with a reagent that detects TSLC1 and detecting a reduction in the TSLC1 as compared to that of a comparable normal cell, preferably in said detection an increased methylation of the *TSLC1* promoter in the test cell and/or a reduced production of TSLC1 in the test cell as compared to the comparable normal cell is determined.

In yet another embodiment, the present invention provides methods of treating high-risk HPV-induced invasive cancers and their precursor lesions associated with modification of TSLC1 production in test cells in a subject in need thereof, said method comprising contacting cells of a patient suffering from said cancer with a therapeutically effective amount of a reagent that increases TSLC1 level in test cells.

In another aspect, the present invention relates to the use of molecular diagnostic markers for the detection of HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1), wherein said marker indicates *TSLC1* promoter methylation, expression of mRNA associated with production of TSLC1 polypeptide and/or allelic loss of the 11q23 chromosome. By such use, the risk of progression to invasive cancer may be determined.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the *TSLC1* 5' regulatory region (-894 to -1) and coding and transcribed 3' non-coding sequences (1 to 1456), derived from Genbank accession codes AP_003172 and NM_014333, respectively. The start and stop codons are represented in italics and are underlined

DETAILED DESCRIPTION OF THE INVENTION

"Expression" refers to the transcription of a gene into structural RNA (rRNA, tRNA) or messenger RNA (mRNA) and, if applicable, subsequent translation into a protein.

The term "HPV-induced invasive cancer" refers to a carcinoma induced by high-risk HPV, which invades surrounding tissue.

The term "invasive cervical cancer" refers to a cervical carcinoma invading surrounding tissue.

The terms "pre-malignant lesion" and "precursor lesion" refer to a stage in the multistep cellular evolution to cancer with a strongly increased chance to progress to a carcinoma. With classical morphology the pathologist is unable to predict in the individual patient which of these lesions will progress or regress. The current invention refers to a method, which can predict the progression to invasive cancer.

The term "invasive potential" refers to the potential to invade surrounding tissue and consequently to become malignant.

The term "pre-malignant cervical lesion" refers to a stage in the multistep cellular evolution to cervical cancer with a strongly increased chance to progress to a cervical carcinoma. With classical morphology the pathologist is unable to predict in the individual patient which of these lesions will progress or regress.

The term "capable of specifically hybridizing to" refers to a nucleic acid sequence capable of specific base-pairing with a complementary nucleic acid sequence and binding thereto to form a nucleic acid duplex.

A "complement" or "complementary sequence" is a sequence of nucleotides which forms a hydrogen-bonded duplex with another sequence of nucleotides according to Watson-Crick base-pairing rules. For example, the complementary base sequence for 5'-AAGGCT-3' is 3'-TTCCGA-5'.

The term "stringent hybridization conditions" refers to hybridization conditions that affect the stability of hybrids, e.g., temperature, salt concentration, pH, formamide concentration and the like. These conditions are empirically optimised to maximize specific binding and minimize non-specific binding of the primer or the probe to its target nucleic acid sequence. The terms as used include reference to conditions under which a probe or primer will hybridise to its target sequence, to a detectably greater degree than other sequences (e.g. at least 2-fold over background). Stringent conditions are sequence dependent and will be different in different circumstances. Longer sequences hybridise specifically at higher temperatures. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridises to a perfectly matched probe or primer. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes or primers (e.g. 10 to 50 nucleotides) and at least about 60°C for long probes or primers (e.g. greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringent conditions or "conditions of reduced stringency" include hybridization with a buffer solution of 30% formamide, 1 M NaCl, 1% SDS at 37°C and a wash in 2x SSC at 40°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1x SSC at 60°C. Hybridization procedures are well known in the art and are described in e.g. Ausubel et al, Current Protocols in Molecular Biology, John Wiley & Sons Inc., 1994.

The term "oligonucleotide" refers to a short sequence of nucleotide monomers (usually 6 to 100 nucleotides) joined by phosphorous linkages (e.g.,

phosphodiester, alkyl and aryl-phosphate, phosphorothioate), or non-phosphorous linkages (e.g., peptide, sulfamate and others). An oligonucleotide may contain modified nucleotides having modified bases (e.g., 5-methyl cytosine) and modified sugar groups (e.g., 2'-O-methyl ribosyl, 2'-O-methoxyethyl ribosyl, 2'-fluoro ribosyl, 2'-amino ribosyl, and the like).

Oligonucleotides may be naturally-occurring or synthetic molecules of double- and single-stranded DNA and double- and single-stranded RNA with circular, branched or linear shapes and optionally including domains capable of forming stable secondary structures (e.g., stem-and-loop and loop-stem-loop structures).

The term "primer" as used herein refers to an oligonucleotide which is capable of annealing to the amplification target allowing a DNA polymerase to attach thereby serving as a point of initiation of DNA synthesis when placed under conditions in which synthesis of primer extension product which is complementary to a nucleic acid strand is induced, i.e., in the presence of nucleotides and an agent for polymerization such as DNA polymerase and at a suitable temperature and pH. The (amplification) primer is preferably single stranded for maximum efficiency in amplification. Preferably, the primer is an oligodeoxy ribonucleotide. The primer must be sufficiently long to prime the synthesis of extension products in the presence of the agent for polymerization. The exact lengths of the primers will depend on many factors, including temperature and source of primer. A "pair of bi-directional primers" as used herein refers to one forward and one reverse primer as commonly used in the art of DNA amplification such as in PCR amplification.

The term "probe" refers to a single-stranded oligonucleotide sequence that will recognize and form a hydrogen-bonded duplex with a complementary sequence in a target nucleic acid sequence analyte or its cDNA derivative.

The *TSLC1* (tumor suppressor in lung cancer 1) gene (Genbank Accession NM_014333) has originally been identified as a tumor suppressor gene in the lung cancer cell line A549 by functional complementation studies

(Kuramochi et al., 2001; Nature Genet 27, 427-430). It was shown that re-expression of *TSLC1* in the lung cancer cell line A549 suppressed tumorigenicity in nude mice. Moreover, loss or suppression of *TSLC1* expression in other lung cancer cell lines was shown to be correlated to both
5 tumorigenicity and spleen to liver metastasis in nude mice. *TSLC1* mRNA suppression in these cell lines was found to correlate to hypermethylation of the *TSLC1* promoter.

Subsequent analysis of non-small cell lung cancers (NSCLC) revealed allelic loss at 11q23.2, the *TSLC1* locus, in about 40% of lung cancers
10 and within this group of tumors with allelic loss promoter hypermethylation of the other allele could be detected in 85% of cases.

TSLC1 encodes a member of the immunoglobulin super family of cell adhesion molecules (IgCAMs), which consists of a wide variety of cell-
surface molecules that are characterized by an immunoglobulin unit. The
15 *TSLC1* protein is an N-linked glycoprotein of 75 kDa, which is localized at the cell membrane and is involved in intracellular adhesion through homophilic trans-interaction (Masuda et al, 2002; J Biol Chem 277, 31014-31019).

The present inventors have now established that *TSLC1* silencing is a frequent event in cervical cancer cell lines. *TSLC1* silencing is found to result
20 from *TSLC1* promoter hypermethylation either or in not in combination with allelic loss. In vitro studies revealed a functional involvement of *TSLC1* inactivation in both anchorage independent growth and tumorigenicity of cervical cancer cells, whereas immortality and proliferation were not affected. This points to a role for *TSLC1* silencing in tumor invasion rather than
25 proliferation. Loss of a putative tumor suppressor gene on chromosome 11 was already known to be involved in tumorigenicity of SiHa cervical cancer cells, and allelic loss at 11q22-23 has frequently been detected in invasive cervical carcinoma. These results indicate that inactivation of *TSLC1* at 11q23.2 might play a crucial role in cervical cancer invasion.

Analysis of cervical tissue specimens revealed that *TSLC1* promoter hypermethylation is limited to only a subset of high-grade CIN lesions but detectable in up to 58% of invasive cervical carcinomas. Moreover, *TSLC1* promoter hypermethylation can specifically be detected in cervical smears
5 derived from women with cervical cancer. Analysis of cervical cancer cell lines revealed that silencing of *TSLC1* expression is present in as much as 91%
(10/11) of cell lines. *TSLC1* silencing appeared not only associated with promoter methylation, but also with allelic loss at the *TSLC1* locus and yet
unknown events, suggesting that the percentage of cervical cancers showing
10 *TSLC1* promoter methylation is even an underestimation of the actual percentage of cases in which the *TSLC1* gene is silenced.

Accordingly, the present invention provides methods of detecting HPV-induced invasive cancers and their precursor lesions associated with
tumor suppressor lung cancer 1 (*TSLC1*) in a subject in need thereof, or
15 indicative thereof, said method comprising contacting a cell component of a test cell of the subject with a reagent that detects the level of the cell component in the test cell and determining a modification in the level of the cell component in the test cell as compared with a comparable healthy cell,
wherein the cell component indicates the level of *TSLC1* in the cell and the
20 modification indicates the presence of HPV-induced invasive cancers and their precursor lesions.

The test cell of the subject may comprise a cell from a sample of skin cells (e.g. in the case of warts, verrucas and the like, presumably caused by cutaneous HPV infections), a sample of mucosal cells, such as cervical cells,
25 and also other tissue such as oral cavity, oropharynx, , penis, vulva, anus, rectum and other tissues wherein a cancer associated with HPV is to be detected. All such samples may be used as a sample in a method of the present invention. Preferably, a sample of a patient's cells comprise cervical cells as test cells.

A method of the present invention is particularly suited for the detection of HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1) that are induced by high-risk HPVs. A method of detecting HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1) may accordingly relate to the measurement of *TSLC1* expression, such as in the form of measuring *TSLC1* gene transcripts and/or subsequent proteins translated from said transcripts. Also a method of detecting HPV-induced invasive cancers and their precursor lesions may comprise measuring *TSLC1* promoter methylation as an indication of *TSLC1* expression capacity and/or TSLC1 protein production capacity.

Figure 1 shows the cg-rich promoter region upstream of the atg start codon in the *TSLC1* gene and the coding region for the TSLC1 protein. Methylation of the cg-rich promoter region will result in a sharply decreased transcription or even complete blockage of transcription. Therefore, the promoter region provides a positive marker sequence for the expression potential of this gene. Alternatively, the expression of the *TSLC1* gene may be detected by measuring gene transcripts. As such, the coding region for the TSLC1 protein in this gene provides a marker sequence for detection of transcripts of the gene. In yet another alternative, the expression of the *TSLC1* gene may be detected by measuring TSLC1 protein directly.

The test cell component contacted can thus be nucleic acid, such as DNA or RNA, preferably mRNA, or protein. When a cell component is protein, the reagent is typically an anti-TSLC1 antibody. When the component is nucleic acid, the reagent is typically a nucleic acid (DNA or RNA) probe or (PCR) primer. By using such probes or primers, gene expression products, such as mRNA may for example be detected. Alternatively, when the component is nucleic acid, the reagent may also be a restriction endonuclease, preferably a methylation sensitive restriction endonuclease for the detection of the presence

of methyl groups on the test cell nucleic acid, said test cell nucleic acid then preferably being DNA.

The test cell component may be detected directly *in situ* or it may be isolated from other cell components by common methods known to those of skill in the art before contacting with the reagent (see for example, "Current Protocols in Molecular Biology", Ausubel et al. 1995. 4th edition, John Wiley and Sons; "A Laboratory Guide to RNA: Isolation, analysis, and synthesis", Krieg (ed.), 1996, Wiley-Liss; "Molecular Cloning: A laboratory manual", J. Sambrook, E.F. Fritsch. 1989. 3 Vols, 2nd edition, Cold Spring Harbor Laboratory Press)

Detection methods include such analyses as Southern and Northern blot analyses, RNase protection, immunoassays, *in situ* hybridization, PCR (Mullis 1987, U.S. Pat. No. 4,683,195, 4,683,202, en 4,800,159), LCR (Barany 1991, Proc. Natl. Acad. Sci. USA 88:189-193; EP Application No., 320,308), 3SR (Guatelli *et al.*, 1990, Proc. Natl. Acad. Sci. USA 87:1874-1878), SDA (U.S. Pat. Nos. 5,270,184, en 5,455,166), TAS (Kwoh *et al.*, Proc. Natl. Acad. Sci. USA 86:1173-1177), Q-Beta Replicase (Lizardi *et al.*, 1988, Bio/Technology 6:1197), Rolling Circle Amplification (RCA) or other methods for the amplification of DNA. In an alternative method RNA may be detected by such methods as NASBA (L. Malek *et al.*, 1994, Meth. Molec. Biol. 28, Ch. 36, Isaac PG, ed., Humana Press, Inc., Totowa, N.J.) or TMA.

Nucleic acid probes, primers and antibodies can be detectably labeled, for instance, with a radioisotope, a fluorescent compound, a bioluminescent compound, a chemiluminescent compound, a metal chelator, or an enzyme. Those of ordinary skill in the art will know of other suitable labels for binding to the reagents or will be able to ascertain such, using routine experimentation.

Other methods for detection include such analyses as can be performed with nucleic acid arrays (See i.a. Chee *et al.*, 1996, Science 274(5287):610-614). For example, DNA arrays may be used for the detection of

nucleic acids according to the invention. Such arrays comprise oligonucleotides with sequences capable of hybridizing under stringent conditions to the nucleic acid cell component of which the level is detected in a method of the present invention.

5 Since the present invention shows that a decreased level of *TSLC1* transcription is often the result of hypermethylation of the *TSLC1* gene, it is often desirable to directly determine whether the *TSLC1* gene is hypermethylated. In particular, the cytosine rich areas termed "CpG islands", which lie in the 5' regulatory regions of genes are normally unmethylated. The
10 term "hypermethylation" includes any methylation of cytosine at a position that is normally unmethylated in the *TSLC1* gene sequence (e. g. the *TSLC1* promoter). Hypermethylation can for instance be detected by restriction endonuclease treatment of the *TSLC1* polynucleotide (gene) and Southern blot analysis. Therefore, in an invention method wherein the cellular component
15 detected is DNA, restriction endonuclease analysis is preferred to detect hypermethylation of the *TSLC1* gene. Any restriction endonuclease that includes CG as part of its recognition site and that is inhibited when the C is methylated, can be utilized. Methylation sensitive restriction endonucleases such as BssHII, MspI, NotI or HpaII, used alone or in combination, are
20 examples of such endonucleases. Other methylation sensitive restriction endonucleases will be known to those of skill in the art.

Other methods for the detection of *TSLC1* promoter hypermethylation involve bisulfite modification of DNA, in which the unmethylated cytosines are converted to an uracil whereas the methylated
25 cytosines are protected from chemical modification. Subsequent PCR amplification and sequencing will reveal whether cytosines in CpG islands are maintained in case of methylation or replaced by a uracil in case of an unmethylated status. Another method involves the treatment a PCR amplified product generated from bisulfite modified DNA with restriction
30 endonuclease that includes CG as part of its recognition site.

An alternative means to test for methylated sequences is a methylation specific PCR, which is also based on bisulfite modification of DNA, followed by specific PCR reactions that target CpG rich sequences.

For purposes of the invention, an antibody (*i.e.*, an anti-TSLC1 antibody) or nucleic acid probe specific for TSLC1 may be used to detect the presence of TSLC1 polypeptide (using antibody) or *TSLC1* polynucleotide (using nucleic acid probe) in biological fluids or tissues. Oligonucleotide primers based on any coding sequence region and regulatory sequence region in the *TSLC1* sequence are useful for amplifying DNA, for example by PCR.

When using PCR primers, nucleic acid probes or restriction endonucleases, the 5' regulatory region and coding sequence of the *TSLC1* sequence is analysed.

Any specimen containing a detectable amount of *TSLC1* polynucleotide or TSLC1 polypeptide antigen can be used. Nucleic acid can also be analyzed by RNA in situ methods that are known to those of skill in the art such as by *in situ* hybridization. Preferred samples for testing according to methods of the invention include such specimens as (cervical) smears and/or (cervical) biopsies and the like. Preferably, cytologically abnormal (cervical) smears and/or biopsies of high-grade (pre)malignant lesions are used as samples for testing. Although the subject can be any mammal, preferably the subject is human.

The invention methods can utilize antibodies immunoreactive with TSLC1 polypeptide, the predicted amino acid sequence of which is available as GenBank Accession No. BAA75822, or immunoreactive fragments thereof.

Antibody that consists essentially of pooled monoclonal antibodies with different epitopic specificities, as well as distinct monoclonal antibody preparations can be used. Monoclonal antibodies are made from antigen containing fragments of the protein by methods well known to those skilled in the art (Kohler, et al., Nature, 256: 495, 1975).

The term antibody as used in this invention is meant to include intact molecules as well as fragments thereof, such as Fab and F (ab')₂, which are capable of binding an epitopic determinant on TSLC1.

Monoclonal antibodies can be used in the invention diagnostic
5 methods, for example, in immunoassays in which they can be utilized in liquid phase or bound to a solid phase carrier. In addition, the monoclonal antibodies in these immunoassays can be detectably labeled in various ways. Examples of types of immunoassays that can utilize monoclonal antibodies of the invention are competitive and non-competitive immunoassays in either a direct or
10 indirect format. Examples of such immunoassays are the radioimmunoassay (RIA) and the sandwich (immunometric) assay. Detection of the antigens using the monoclonal antibodies of the invention can be done utilizing immunoassays that are run in either the forward, reverse, or simultaneous modes, including
immunohistochemical assays on physiological samples. Those of skill in the art
15 will know, or can readily discern, other immunoassay formats without undue experimentation.

Monoclonal antibodies can be bound to many different carriers and used to detect the presence of TSLC1. Examples of well-known carriers include glass, polystyrene, polypropylene, polyethylene, dextran, nylon, amylases,
20 natural and modified celluloses, polyacrylamides, agaroses and magnetite. The nature of the carrier can be either soluble or insoluble for purposes of the invention. Those skilled in the art will know of other suitable carriers for binding monoclonal antibodies, or will be able to ascertain such using routine experimentation.

25 In performing the assays it may be desirable to include certain "blockers" in the incubation medium (usually added with the labeled soluble antibody). The "blockers" are added to assure that non-specific proteins, proteases, or antiheterophilic immunoglobulins to anti-TSLC1 immunoglobulins present in the experimental sample do not cross-link or
30 destroy the antibodies on the solid phase support, or the radiolabeled indicator

antibody, to yield false positive or false negative results. The selection of "blockers" therefore may add substantially to the specificity of the assays described in the present invention. A number of nonrelevant (i. e., nonspecific) antibodies of the same class or subclass (isotype) as those used in the assays (e. g., IgG1, IgG2a, IgM, etc.) can be used as "blockers". The concentration of the "blockers" (normally 1-100 $\mu\text{g}/\mu\text{L}$) may be important, in order to maintain the proper sensitivity yet inhibit any unwanted interference by mutually occurring cross-reactive proteins in the specimen.

In using a monoclonal antibody for the *in vivo* detection of antigen, the detectably labeled monoclonal antibody is given in a dose that is diagnostically effective. The term "diagnostically effective" means that the amount of detectably labeled monoclonal antibody is administered in sufficient quantity to enable detection of the site having the TSLC1 antigen for which the monoclonal antibodies are specific. The concentration of detectably labeled monoclonal antibody which is administered should be sufficient such that the binding to those cells having TSLC1 is detectable compared to the background, depending upon the *in vivo* imaging or detection method employed, such as MRI, CAT scan, and the like. Further, it is desirable that the detectably labeled monoclonal antibody be rapidly cleared from the circulatory system in order to give the best target-to-background signal ratio.

As a rule, the dosage of detectably labeled monoclonal antibody for *in vivo* diagnosis will vary depending on such factors as age, sex, and extent of disease of the individual. The dosage of monoclonal antibody can vary from about 0.001 mg/m^2 to about 500 mg/m^2 , preferably 0.1 mg/m^2 to about 200 mg/m^2 , most preferably about 0.1 mg/m^2 to about 10 mg/m^2 . Such dosages may vary, for example, depending on whether multiple injections are given, tumor burden, and other factors known to those of skill in the art.

For *in vivo* diagnostic imaging, the type of detection instrument available is a major factor in selecting a given radioisotope. The radioisotope chosen must have a type of decay that is detectable for a given type of

instrument. Still another important factor in selecting a radioisotope for in vivo diagnosis is that the half-life of the radioisotope be long enough so that it is still detectable at the time of maximum uptake by the target, but short enough so that deleterious radiation with respect to the host is minimized.

- 5 Ideally, a radioisotope used for in vivo imaging will lack a particle emission, but produce a large number of photons in the 140-250 keV range, which may be readily detected by conventional gamma cameras.

For *in vivo* diagnosis, radioisotopes can be bound to immunoglobulin either directly or indirectly by using an intermediate functional group.

- 10 Intermediate functional groups which often are used to bind radioisotopes which exist as metallic ions to immunoglobulins are the bifunctional chelating agents such as diethylenetriaminepentaacetic acid (DTPA) and ethylenediaminetetraacetic acid (EDTA) and similar molecules. Typical
 15 examples of metallic ions that can be bound to the monoclonal antibodies of the invention are ^{111}In , ^{97}Ru , ^{67}Ga , ^{68}Ga , ^{72}As , ^{89}Zr , and ^{201}Tl .

- A monoclonal antibody useful in the invention methods can also be labeled with a paramagnetic isotope for purposes of *in vivo* diagnosis, as in magnetic resonance imaging (MRI) or electron spin resonance (ESR). In general, any conventional method for visualizing diagnostic imaging can be
 20 utilized. Usually gamma and positron emitting radioisotopes are used for camera imaging and paramagnetic isotopes for MRI. Elements that are particularly useful in such techniques include ^{157}Gd , ^{55}Mn , ^{162}Dy , ^{52}Cr and ^{56}Fe .

- Other, *e.g. ex vivo*, diagnostic methods for the detection of TSLC1
 25 production, *TSLC1* gene expression or disorders therein, include methods wherein a sample for testing is provided, which sample comprises a cell preparation from cervical or other tissue. Preferably such samples are provided as smears. In order to provide for efficient testing schemes, cytologically abnormal (cervical) smears and/or biopsies of high-grade
 30 (pre)malignant lesions are used as samples for testing.

A tissue sample obtained from a mammal, preferably a human, is suitably pretreated to allow contact between a target cellular component of a test cell comprised in said sample with a reagent that detects TSLC1 and detecting a reduction in the TSLC1 as compared to that of a comparable normal cell. Samples may be mounted on a suitable support to allow observation of individual cells. Examples of well-known support materials include glass, polystyrene, polypropylene, polyethylene, polycarbonate, polyurethane, optionally provided with layers to improve cell adhesion and immobilization of the sample, such as layers of poly-L-lysine or silane. Cervical smears or biopsies may for instance be prepared as for the Papanicolaou (Pap) test or any suitable modification thereof as known by the skilled person, and may be fixed by procedures that allow proper access of the reagent to the target component. If storage is required, routine procedures use buffered formalin for fixation followed by paraffin embedding, which provides for a well-preserved tissue infrastructure. In order to allow for immunohistochemical or immunofluorescent staining, the antigenicity of the sample material must be retrieved or unmasked. One method of retrieving the antigenicity of formaldehyde cross-linked proteins involves the treatment of the sample with proteolytic enzymes. This method results in a (partial) digest of the material and mere fragments of the original proteins can be accessed by antibodies.

Another method for retrieving the immunoreactivity of formaldehyde cross-linked antigens involves the thermal processing using heat or high energy treatment of the samples. Such a method is described in e.g. U.S. Pat. No. 5,244,787. Yet another method for retrieving antigens from formaldehyde-fixed tissues is the use of a pressure cooker, either in combination with a microwave or in the form of an autoclave, such as described in e.g. Norton, 1994, *J. Pathol.* 173(4):371-9 and Taylor et al. 1996, *Biotech Histochem* 71(5):263-70.

Several alternatives to formaldehyde may be used, such as ethanol, methanol, methacarn or glyoxal, citrated acetone, or fixatives may be used in

combination. Alternatively, the sample may be air-dried before further processing.

In order to allow for a detection with nucleic acid probes, the sample material must be retrieved or unmasked in case of formalin fixed and paraffin embedded material. One method involves the treatment with proteolytic enzymes and a postfixation with paraformaldehyde. Proteolytic digestion may be preceded by a denaturation step in HCl. This method results in a (partial) digest of the material allowing the entry of probes to the target. No specific unmasking procedures are required in case of non-formalin fixed material, e.g. frozen material. Prior to hybridisation samples can be acetylated by treatment with triethanolamine buffer.

The nucleic acid probes or antibodies are then contacted with the sample material in a suitable buffer and permitted to specifically hybridize or bind to their nucleic acid or protein target. Upon specific binding of the nucleic acid probes or antibodies to the target components, labeled probes and/or antibodies may be detected by such methods as confocal laser scanning microscopy, bright field microscopy, flow cytometry optionally in combination with fluorescence associated cell sorting, or modifications of these techniques, which are well known to the person skilled in the art.

In one embodiment of a method of the invention an increased methylation of the *TSLC1* promoter in the test cell and/or reduced production of *TSLC1* in the test cell is detected as compared to the comparable normal cell.

The present invention also provides methods for treating a subject with HPV-induced invasive cancers associated with modification of *TSLC1* production, or indicative thereof, comprising administering to a subject with the cancer a therapeutically effective amount of a reagent that increases *TSLC1* expression. In HPV-induced invasive cancers associated with tumor suppressor lung cancer 1 (*TSLC1*), the *TSLC1* nucleotide sequence is underexpressed as compared to expression in a normal cell, therefore, it is

possible to design appropriate therapeutic or diagnostic techniques directed to this sequence. Thus, nucleic acid sequences that increase *TSLC1* expression at the transcriptional or translational level can be used and, for example, nucleic acid sequences encoding *TSLC1* (sense) could be administered to the subject
 5 with the HPV-induced invasive cancer, such as invasive cervical cancer.

The term "invasive cervical cancer" denotes malignant as well as premalignant cell populations, which often appear to differ from the surrounding tissue both morphologically and genotypically. These disorders are found to be associated with absence of or reduced expression of *TSLC1*.
 10 Essentially, any test cell abnormality that is etiologically linked to expression of *TSLC1* could be considered susceptible to treatment using methods of the present invention that employ a reagent to increase *TSLC1* expression.

Increased *TSLC1* expression may be attained by suppression of methylation of *TSLC1* polynucleotide when *TSLC1* is under-expressed. When,
 15 upon diagnosis according to the present invention, the HPV-induced cancer detected is associated with *TSLC1* expression, such methylation suppressive reagents as 5-azacytadine can be introduced to a cell. Alternatively, when, upon diagnosis according to the present invention, the invasive cervical cancer detected is associated with under-expression of *TSLC1* polypeptide, a sense
 20 polynucleotide sequence (the DNA coding strand) encoding *TSLC1* polypeptide, or 5' regulatory nucleotide sequences (i. e., promoter) of *TSLC1* in operable linkage with *TSLC1* polynucleotide can be introduced into the cell. Demethylases known in the art could also be used to remove methylation.

The present invention also provides gene therapy for the treatment
 25 of HPV-induced cancer associated with modification of *TSLC1* production. Such therapy would achieve its therapeutic effect by introduction of the appropriate *TSLC1* polynucleotide that contains a *TSLC1* structural gene (sense), into cells of subjects having HPV-induced cancer. Schemes and procedure for effecting gene therapeutic treatment are known in the art, e.g. in
 30 WO 02/14557. Delivery of sense *TSLC1* polynucleotide constructs can be

achieved by using an expression vector or by using a colloidal dispersion system, preferably a recombinant expression vector, said recombinant expression vector preferably being a plasmid, a viral particle or a phage.

The polynucleotide sequences used in the methods of the invention
5 may be the native, unmethylated sequence or, alternatively, may be a sequence in which a nonmethylatable analog is substituted within the sequence. Preferably, the analog is a nonmethylatable analog of cytidine, such as 5-azacytadine. Other analogs will be known to those of skill in the art. Alternatively, such nonmethylatable analogs could be administered to a
10 subject as drug therapy, alone or simultaneously with a sense structural gene for *TSLC1* or sense promoter for *TSLC1* operably linked to *TSLC1* structural gene. Preferably the *TSLC1* polynucleotide used in the invention methods is derived from a mammalian organism, and most preferably from human.

Allelic loss of the *TSLC1* locus (or LOH) can be detected by PCR
15 amplification of polymorphic sequences flanking the *TSLC1* gene. Both DNA from test cells and from healthy cells of the same individual are PCR amplified. The two PCR fragments representing both alleles are separated on a polyacrylamide gel or by capillary electrophoresis. PCR products derived from DNA of a healthy cells and of test cells are compared, wherein loss of one of
20 two PCR fragments in test cells is indicative of an allelic loss/genetic deletion of *TSLC1* locus.

The present invention also provides a kit of parts for use in a method of detecting HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (*TSLC1*) in test cells of a
25 subject. Such a kit may suitably comprise a brush or spatula to take a (cervical) scrape together with a container filled with collection medium to collect test cells. Alternatively, a sampling device consisting of an irrigation syringe, a disposable female urine catheter and a container with irrigation fluid will be included to collect cervical cells by cervico-vaginal lavage.

A kit according to the present invention may comprise primers and probes for the detection of *TSLC1* promoter methylation, for the detection of allele losses at chromosome 11q23.2 or for the detection of *TSLC1* mRNA expression. In another embodiment, a kit according to the invention may
5 comprise antibodies and reagents for the detection of *TSLC1* protein expression in cervical scrapes or tissue specimens.

A kit of parts according to the invention comprises means for the detection of *TSLC1* promoter methylation or *TSLC1* expression, such as *TSLC1*-specific antibodies, methylation-sensitive restriction enzymes, or
10 probes or primers capable of hybridising to the nucleotide sequence of Fig. 1.

In yet another alternative embodiment of a kit of the invention the means for the detection of *TSLC1* promoter methylation or *TSLC1* expression may be combined with means for the detection of HPV infection, preferably for the detection of HPV infection of the high-risk type. Such means may comprise
15 HPV-specific primers or probes as are known in the art.

The present invention will now be illustrated by way of the following, non limiting examples.

EXAMPLES

20 Example 1a. Frequent *TSLC1* silencing in cervical carcinoma cell lines

TSLC1 mRNA expression levels were measured in 11 cervical carcinoma cell lines by real time quantitative RT-PCR using Lightcycler technology (Roche). Compared to normal primary epithelial cells, *TSLC1* mRNA was undetectable in 8 of the 11 cell lines and severely reduced in
25 another 2 cell lines, showing that *TSLC1* downregulation is apparent in 91% (10/11) of cervical carcinoma cell lines analysed.

On the other hand, *TSLC1* expression was still abundant in HPV-immortalized cells. These HPV-immortalized cells are not yet tumorigenic and have previously been shown to be representative of premalignant cervical
30 lesions in vivo.

Subsequently, the analysis of the mechanisms underlying *TSLC1* downregulation was undertaken. Besides genetic events, i.e. deletions and inactivating mutations, epigenetic events may result in gene silencing, as has been described for lung cancers [Kuramochi et al., 2001].

5

Example 1b. Role of methylation in *TSLC1* silencing in cervical cancer cells

To assess whether a methylating event was underlying *TSLC1* silencing in cervical cancer cells, the cell lines SiHa, HeLa and CaSki were treated with the methylation inhibitor 5-aza-cytidine. *TSLC1* mRNA levels were compared to expression levels in primary epithelial cells, which was set to a 100%. Upon 5 to 7 days of incubation with 5-azacytidine *TSLC1* expression levels were upregulated from 0% to 26% in SiHa, and from 4% to 70% and 0% to 33% in HeLa and CaSki cells, respectively. These data indicate that in all these 3 cell lines *TSLC1* downregulation results, at least in part, from a methylating event.

10

15

Next, *TSLC1* promoter methylation by radioactive bisulfite sequencing was studied. It was found that all 6 CpG sites sequenced were methylated in 9/11 cervical cancer cell lines. Except for one cell line, no wild type sequences were detected in these cell lines, indicating that *TSLC1* promoter methylation was clonal. In the remaining 2 cell lines none of the CpG sites were methylated. Except for one cell line, *TSLC1* promoter methylation was correlated to reduced or undetectable *TSLC1* mRNA expression. No *TSLC1* promoter methylation was detected in 4 isolates of normal epithelial cells and in the non-tumorigenic HPV-immortalized cells.

20

25

Example 1c. Role of a chromosomal deletion in *TSLC1* silencing

To analyse whether a chromosomal deletion attributed to *TSLC1* silencing loss of heterozygosity (LOH) analysis was performed using 8 polymorphic markers flanking the *TSLC1* gene. Normal DNA derived from either lymphoblasts or fibroblasts of 8 cervical carcinoma cell lines was

30

available. In 3 (38%) of these cell lines an allelic loss at 11q23.2 was apparent. All 3 cell lines revealed *TSLC1* promoter hypermethylation and absence of detectable *TSLC1* mRNA, suggesting that allelic deletion combined with promoter hypermethylation underlied complete gene silencing.

5 Analysis on the HPV immortalized cell lines showed an LOH at 11q23.2 in all immortal passages of one of the four cell lines. However, *TSLC1* expression was still detectable in these passages and no promoter methylation was found, indicating that the retained allele is still actively transcribed. No allelic loss at 11q23 was detected in the other three cell lines.

10 Taken together, these data show that *TSLC1* silencing in cervical cancer cells may result from 1) promoter methylation of one allele combined with deletion of the other allele as found in 3/8 cell lines, 2) promoter methylation without an allelic loss, suggesting that either both alleles are hypermethylated or that one is mutated, as was found in two cell lines, or 3) 15 other yet unknown mechanisms, as was found in two cell lines.

Example 2. *TSLC1* suppresses anchorage independent growth in vitro and tumorigenicity in nude mice

20 Malignant transformation of hr-HPV infected epithelial cells in vitro has been shown to proceed via the subsequent acquisition of 1) an immortal phenotype and 2) an anchorage independent phenotype, which can be measured by the growth of cells in soft agarose.

To test whether *TSLC1* silencing is functionally involved in the development of cervical cancer, the *TSLC1* coding sequence was transfected in 25 the cervical cancer cell line SiHa in which endogenous *TSLC1* expression was undetectable. Re-expression of *TSLC1* in these *TSLC1* mRNA negative SiHa cells resulted in a highly reduced growth in soft agarose, which is a measure for anchorage independence, whereas the growth in normal monolayer was unaffected. Moreover, re-expression of *TSLC1* in SiHa cells resulted in a 30 suppression of tumor growth in nude mice in two of four SiHa/*TSLC1*

transfectants and tumors that did arise from the SiHa/TSLC1 transfectants grew slower than the controls.

In conclusion, these results show that TSLC1 can suppress both anchorage independent and tumorigenic growth of cervical cancer cells, leaving immortality and proliferation unaffected.

Example 3.-Silencing of TSLC1 in cervical tissue specimens.

To determine whether and at what stage during cervical carcinogenesis *TSLC1* silencing occurs *in vivo*, the methylation status of the *TSLC1* promoter was analyzed in cervical tissue specimens.

Since the tissue specimens consist of an admixture of normal stromal components and abnormal cells the sensitivity of the assay to detect methylated CpG islands in a background of normal unmethylated DNA was determined. By analysing a dilution series of SiHa DNA (methylated *TSLC1* promoter) in DNA derived from primary keratinocytes (unmethylated *TSLC1* promoter), it was found that as low as 5% of methylated DNA in a background of unmethylated DNA could still be detected with high reliability, using radioactive bisulfite sequencing on a Genomymx device.

Using this method none of 15 normal cervical epithelial biopsies revealed *TSLC1* promoter methylation. Similarly, *TSLC1* promoter methylation was undetectable in all low grade CIN lesions (n=10). Of 30 high grade CIN lesions 35% showed methylation of the *TSLC1* promoter. In addition, a total of 50 cervical squamous cell carcinoma sections were analysed, 58% (29/50) of which showed *TSLC1* promoter methylation. Thus, *TSLC1* promoter methylation appears to be a rather frequent event in cervical squamous cell carcinomas and occurs late during the multistep sequence of carcinogenesis. Since in addition to *TSLC1* promoter hypermethylation other alterations, including an allelic loss at 11q23.2, may contribute to silencing of *TSLC1* the actual percentage of *TSLC1* silencing in cervical carcinomas is likely to be even higher than 58%.

Example 4. Detection of *TSLC1* promoter methylation in cervical smears

In a small pilot study it was assessed whether the detection of *TSLC1* alterations can be applied to cervical smears and as such be used as a marker in cervical cancer screening programs.

For this, *TSLC1* promoter methylation was analysed in archival smears of women who developed cervical cancer. These scrapes were derived from a retrospective case-control which was designed to determine the value of hr-HPV testing to signal false negative cervical smears in women who developed cervical cancer and to assess whether hr-HPV is present in normal smears preceding cervical cancer (Zielinski et al., 2001; Br J Cancer 85, 398-404). Analysis of 9 index smears, i.e. smears taken at the time of cervical cancer diagnosis and classified as Pap4 or Pap5 (i.e. suspect for carcinoma in situ/invasive cancer), as well as the 9 associated cervical carcinoma biopsies, showed that *TSLC1* promoter methylation could be detected in smears of women with a *TSLC1* promoter methylation positive cancer biopsy. No *TSLC1* promoter methylation was detected in control smears (n=12).

As a follow up it was determined whether the detection of *TSLC1* promoter methylation cannot only provide a diagnostic marker to detect cervical cancer but may also provide a marker for risk assessment of premalignant cervical lesions. Hereto, the same patient group was studied as described above and archival cervical smears were analysed which were taken 1-2 years before cervical cancer was diagnosed and which were classified as Pap3a2/3b, i.e. moderate/severe dyskaryosis. In one patient, *TSLC1* promoter methylation could already be detected in a cancer preceding smear, which was taken 1 year prior to cervical cancer diagnosis.

31. 03. 2003

(83)

Claims

1. A method of detecting an HPV-induced invasive cancer or precursor lesion thereof associated with tumor suppressor lung cancer 1 (TSLC1) in a subject in need thereof, said method comprising contacting a cell component of a test cell of the subject with a reagent that detects the level of the cell component in the test cell and determining a modification in the level of the cell component in the test cell as compared with a comparable healthy cell, wherein the cell component indicates the level of TSLC1 in the cell and the modification indicates the presence of an HPV-induced invasive cancer or precursor lesion thereof.
2. Method according to claim 1, wherein said HPV-induced invasive cancer or precursor lesion thereof is invasive cervical cancer or a premalignant cervical lesion with invasive potential.
3. Method according to claim 1 or 2, wherein said HPV-induced invasive cancer is a high-risk HPV-induced invasive cancer.
4. Method according to any one of the preceding claims, wherein the modification is a decrease in the level of TSLC1.
5. Method according to any one of the preceding claims, wherein the cell component is a nucleic acid associated with production of TSLC1 polypeptide and the reagent targets the nucleic acid in the test cell, said nucleic acid preferably encoding the TSLC1 and regulatory regions.
6. Method according to claims 5, wherein the nucleic acid is RNA, preferably mRNA.
7. Method according to any one of the preceding claims, wherein the reagent is a restriction endonuclease, preferably a methylation sensitive restriction endonuclease.

8. Method according to claim 5 or 6, wherein the reagent is a nucleic acid probe or primer that binds to the nucleic acid, said nucleic acid probe or primer preferably having a detectable label.
9. Method according to claim 8, wherein the nucleic acid probe has a
5 nucleotide sequence selected from the group consisting of:
- a) a polynucleotide sequence capable of hybridizing under stringent conditions to the 5' regulatory region or the coding region of the *TSLC1* sequence as set forth in Figure 1;
 - b) a polynucleotide having at least 70% identity to the
10 polynucleotide of a);
 - c) a polynucleotide complementary to the polynucleotide of a); and
 - d) a polynucleotide comprising at least 15 bases of a nucleotide of a) or b).
-
10. Method according to any one of the claims 1-4, wherein the cell
15 component is a polypeptide and the reagent targets the polypeptide in the test cell, said polypeptide preferably being *TSLC1* and said reagent preferably being an anti-*TSLC1* antibody.
11. Method according to any one of the claims 1-5, wherein said detecting evaluates the methylation status of the *TSLC1* promoter.
- 20 12. A method of detecting HPV-induced invasive cancer or precursor lesion thereof associated with tumor suppressor lung cancer 1 (*TSLC1*) in a subject in need thereof, said method comprising contacting a target cellular component of a test cell with a reagent that detects *TSLC1* and detecting a reduction in the *TSLC1* as compared to that of a comparable normal cell,
25 preferably in said detection an increased methylation of the *TSLC1* promoter in the test cell and/or a reduced production of *TSLC1* in the test cell as compared to the comparable normal cell is determined.
13. Method according to claim 12, wherein the target cellular component is nucleic acid.

14. Method according to claim 13, wherein the nucleic acid is RNA, preferably mRNA.
15. Method according to claim 12, wherein the target cellular component is a protein.
- 5 16. Method according to claim 13 or 14, wherein the reagent is a nucleic acid probe or primer that binds to TSLC1.
17. Method according to claim 15, wherein the reagent is an anti-TSLC1 antibody.
18. Method according to any one of the claims 12-17, wherein the subject
10 has loss of heterozygosity at chromosome 11q23.
19. A method of treating HPV-induced invasive cancers and their precursor lesions associated with modification of TSLC1 production in cells in a subject in need thereof, said method comprising contacting said cells of a
15 patient suffering from said cancer with a therapeutically effective amount of a reagent that increases TSLC1 level in said cells.
20. Method according to claim 19, wherein the reagent is a polynucleotide sequence comprising a TSLC1 sense polynucleotide sequence, preferably said polynucleotide is the native, unmethylated TSLC1 sense sequence.
- 20 21. Method according to claim 20, wherein a nonmethylatable analog is substituted for cytidine within the TSLC1 sense sequence, said nonmethylatable analog preferably being is 5-azacytadine.
22. Method according to claim 20 or 21, wherein said polynucleotide sequence is contained in an expression vector, said expression vector
25 preferably being a plasmid, a viral particle or a phage.
23. Use of a molecular diagnostic marker for the detection of HPV-
induced invasive cancers and their precursor lesions associated with tumor
suppressor lung cancer 1 (TSLC1), wherein said marker indicates *TSLC1*
promoter methylation, expression of mRNA associated with production of
30 TSLC1 polypeptide and/or allelic loss of chromosome 11q23.

24. Kit of parts for use in a method of detecting HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1) in test cells of a subject, said kit comprising means to collect cervical cells and means for the detection of *TSLC1* promoter methylation or
5 *TSLC1* expression.
25. Kit of parts for use in a method of detecting HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1) in test cells of a subject, said kit comprising means for the detection of *TSLC1* promoter methylation or *TSLC1* expression and means for
10 the detection of HPV infection.

31. 03. 2003

(83)

Title: Detection of HPV-induced invasive cancers and their precursor lesions with invasive potential

Abstract

The invention is in the field of medicine and is concerned with a molecular diagnostic marker for HPV-induced invasive cancers and their precursor lesions with invasive potential. In particular the present invention relates to the use of the TSLC1 gene as marker for invasive cervical cancer. The invention provides methods for detecting HPV-induced invasive cancers and their precursor lesions associated with tumor suppressor lung cancer 1 (TSLC1), comprising contacting a target cellular component of a test cell with a reagent that detects TSLC1 and detecting a reduction in the TSLC1 as compared to that of a comparable normal cell. The invention also provides molecular diagnostic markers for premalignant cervical lesions with invasive potential associated with tumor suppressor lung cancer 1 (TSLC1) in cytologically abnormal cervical smears and/or biopsies.

31. 03. 2003

1/1

(83)

-894	ccgctcttca	cctgaagcct	tgactaattt	tttccgttgt	tgtgtaatct
-844	taaatatcta	atattacaaa	tatttcacac	atatattcaa	cacacaccta
-794	tatattaaaa	ccagggagga	gaccctcgac	aagcggagga	gcctgagcat
-744	accctcctcg	atctaccttt	cccagagattc	tgccgcaaaa	agaccgactg
-694	gaaaatctca	gaacccgact	ctacggctgc	cttctccaac	tatccccgag
-644	tctaccgcta	ggctgttgag	cgggctctcc	cgctccgccc	gacgtgcaaa
-594	gcacgcatgc	acttctccca	gattgttttg	tcaatccggg	gacctgcctt
-544	cttactctcc	actcccgcac	agcccccggt	cccaaagatc	tattccttcg
-494	gtgcaagggtg	agtgaaggaa	atttgcaacg	tctgggttcgc	taggccagat
-444	gcactcgggtg	tgccgggacag	aggacctct	taagggagat	tctccagtcg
-394	tcggtntgat	acagcgattg	ctataacatt	ccttaataaa	ggtgtacaag
-344	aagctagacc	cgccccctgg	agccccgagtc	cttgccagcc	agggcgcccg
-294	gagaacattt	ttccttgatc	cggggaaagc	aaaacccgaa	ttttaacata
-244	aacatatattg	catacgccct	tccccttggc	cccgcctta	ggtggcgccg
-194	gcgcgcgcgc	gaacgccagc	gccagggggc	ggggtggggg	agggagcgag
-144	gccctccgag	agccgggttg	ggctcgcggc	gctgtgattg	gtctgcccg
-94	actccgcctc	cagcgcatgt	cattagcatc	tcattagctg	tccgctcggg
-44	ctccggaggc	agccaacgcc	gccagtctga	ggcaggtgcc	cgacatggcg
+7	agtgtagtgc	tgccgagcgg	atcccagtg	gcggcggcag	cggcggcggc
+57	ggcgctccc	gggctccggc	tccggcttct	gctgttgctc	ttctccgccg
+107	ggcactgatc	cccacaggtg	atgggcagaa	tctgtttacg	aaagacgtga
+157	cagtgatcga	gggagagggt	gcgaccatca	gttgccaagt	caataagagt
+207	gacgactctg	tgattcagct	actgaatccc	aacaggcaga	ccatttatatt
+257	cagggacttc	aggcctttga	aggacagcag	gtttcagttg	ctgaattttt
+307	ctagcagtga	actcaaagta	tcattgacaa	acgtctcaat	ttctgatgaa
+357	ggaagatact	tttgccagct	ctataccgat	ccccacagg	aaagttacac
+407	caccatcaca	gtcctggtcc	caccacgtaa	tctgatgatc	gatataccaga
+457	gagacactgc	ggtggaagg	gaggagattg	aagtcaactg	cactgctatg
+507	gccagcaagc	cagccacgac	tatcaggtgg	ttcaaaggga	acacagagct
+557	aaaaggcaaa	tcggaggtgg	aagagtgggtc	agacatgtac	actgtgacca
+607	gtcagctgat	gctgaagggtg	cacaaggagg	acgatgggg	cccagtgatc
+657	tgccaggtgg	agcaccctgc	ggtcactgga	aacctgcaga	cccagcggtg
+707	tctagaagta	cagtataagc	cacaagtgc	cattcagatg	acttatcctc
+757	tacaaggctt	aacccgggaa	ggggacgcgc	ttgagttaac	atgtgaagcc
+807	atcgggaagc	cccagcctgt	gatggtaact	tgggtgagag	tcgatgatga
+857	aatgcctcaa	cacgcctgac	tgtctgggccc	caacctgttc	atcaataacc
+907	taaacaaaac	agataatggt	acataccgct	gtgaagcttc	aaacatagtg
+957	gggaaagctc	actcggatta	tatgctgtat	gtatacgatc	ccccacaaac
+1007	tatccctcct	cccacaacaa	ccaccaccac	caccaccacc	accaccacca
+1057	ccatccttac	catcatcaca	gattcccagag	caggtgaaga	aggctcgatc
+1107	agggcagtg	atcatgccgt	gatcgggtggc	gtcgtggcgg	tggtggtgtt
+1157	cgccatgctg	tgcttgctca	tcattctggg	gcgctatatt	gccagacata
+1207	aaggtacata	cttcaactcat	gaagccaaag	gagccgatga	cgcagcagac
+1257	gcagacacag	ctataatcaa	tgacagaagga	ggacagaaca	actccgaaga
+1307	aaagaaagag	tacttcatct	agatcagcct	ttttgtttca	atgaggtgtc
+1357	caactggccc	tatttagatg	ataaagagac	agtgatattg	gaacttgcca
+1407	gaaattcgtg	tgttttttta	tgaatgggtg	gaaaggtgtg	agactgggaa

Figure 1

